Synthesis of 7-Deoxy- and 7,10-Dideoxytaxol via Radical Intermediates

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Summary: 7-Deoxytaxol and 7,10-dideoxytaxol were prepared from baccatin III, employing the Barton deoxygenation procedure and the Holton acylation method at C-13.

The important anticancer agent taxol,¹ recently approved for the treatment of ovarian cancer, has been the target of several structure-activity studies.² We have recently shown that, while the benzoate moiety at C-2 of the taxol core is essential for its biological activity,3 the acetate group at C-10 contributes very little to receptor binding.⁴ In this paper we wish to report the synthesis of another deoxygenated taxol analog, 7-deoxytaxol, together with its congener, 7,10-dideoxytaxol. Our studies confirm that the C-7 hydroxyl group is also not essential for antitumor activity.



Our approach is illustrated in Scheme I. Treatment of baccatin with carbon disulfide and sodium hydride in THF produced xanthate 2 in modest yield. This compound was too insoluble in hydrocarbon solvents and had to be silvlated at C-13 prior to deoxygenation. Tin hydride reduction and desilvlation at C-13 afforded 7-deoxybaccatin 3. Acylation according to the Holton protocol⁵ gave the desired 4.

Since deoxygenation at C-10 is not accompanied by loss of bioactivity, we were interested in examining the effect of defunctionalization at both the C-7 and C-10 position on the antitumor activity.

The known 7-(triethylsilyl)baccatin 6 (Scheme II)⁶ was functionalized to give thionocarbonate 7, which was reduced with tin hydride in high yield. Xanthate introduction at C-7 was then followed by treatment with tin hydride. Upon heating in toluene, two compounds were cleanly produced. Separation was difficult but was

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^a Conditions: (i) NaH (1.5 equiv), THF/CS₂ (5:1), imidazole (0.05 equiv), 1.5 h, rt, then MeI (3 equiv), rt, 40 min (57%); (ii) EtaSiCl. imidazole, DMF, rt (74%); Bu₃SnH, PhMe, 110 °C (83%); then TBAF (2 equiv), THF, rt (73%); (iii) BuLi (1.5 equiv), THF, -40 °C, then 5 (1.5 equiv), 0 °C, 45 min; then HCl, CH₃CN, -5 °C, 15 min (53%) overall).

achieved by semipreparative HPLC. Along with the expected dideoxy compound 10, which could be converted in low yield to the desired 12 (Scheme II), we obtained an isomer featuring a ¹H-NMR spectrum that was completely different from the ones typical of taxanes. Extensive NMR analysis⁷ established that the isomer had structure 11.

The formation of this isomer can be rationalized by invoking a cascade of radical rearrangements. The initially formed radical, 13 (Scheme III), is a β -keto radical⁸ and can isomerize via alkoxy radical 14 to the isomeric 15.

This places the radical-bearing carbon in close vicinity with the C11,C12 double bond, and 5-exo cyclization⁹ to 16 occurs. This radical is not quenched by the tin hydride but undergoes a remote hydrogen atom transfer from C-3, as in our recently published photochemical rearrangement.¹⁰ The resulting 17 apparently undergoes a novel

⁽¹⁾ Review: Kingston, D. G. I. Pharm. Ther. 1991, 52, 1.

⁽²⁾ For recent, very interesting work, see: Swindell, C. S.; Krauss, N. E.; Horwitz, S. B.; Ringel, I. J. Med. Chem. 1991, 34, 1176. Also: Guéritte-Voegelein, F.; Guénard, D.; Lavelle, F.; Le Goff, M. T.; Mangatal, L.; Potier, P. J. Med. Chem. 1991, 34, 992.

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⁽⁵⁾ Holton, R. A. Presented at the 203rd Meeting of the American Chemical Society, San Francisco, 1991; Abstract ORGN 0355. See also: Ojima, I.; Habus, I.; Zhao, M.; Zucco, M.; Park, Y. H.; Sun, C. M.; Brigaud, T. Tetrahedron 1992, 48, 6985 and references cited therein.

⁽⁷⁾ Connectivity was established by standard H-H and C-H correlation. The stereochemistry at C-8 and C-12 was demonstrated by NOE experiments. Especially diagnostic is the singlet in the proton spectrum for H-2 (indicating H-3 is missing), the olefinic nature of C-4 and C-3 (δ 143.0 and 126.2, respectively), and three-bond couplings between H-19 and C-11 and between H-12 and C-8. COSY and NOE data are shown in the supplementary material.

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⁽⁹⁾ Bothmi, H. D.; Diggan, T. J. S. Overn. Both, Termin Prais 2, 1992, 1777 and references cited therein.
(9) Griller, D.; Ingold, K. U. Acc. Chem. Res. 1980, 13, 317.
(10) Chen, S. H.; Combs, C. M.; Hill, S. E.; Farina, V.; Doyle, T. W. Tetrahedron Lett. 1992, 33, 7679.



^a Conditions: (i) BuLi (1.25 equiv), THF, then $C_{6}F_{6}CSCl$, -20 °C, 90 min (74%); (ii) Bu₃SnH (1.5 equiv), AIBN, PhMe, 90 °C, 1 h (99%); then HCl, CH₃CN, -10 °C, 30 min (76%); (iii) NaH (1.5 equiv), THF/CS₂ (4:1), rt, 2 h, then MeI (3 equiv), rt, 16 h (53%); (iv) Bu₃SnH (5 equiv), AIBN, PhH, 80 °C, 3h (56% yield of 10, 22% yield of 11); (v) BuLi (1.15 equiv), THF, -40 °C, then 5 (2 equiv), 0 °C, 45 min; then TBAF, THF, 0 °C (30% overall).

oxetane fragmentation reaction,¹¹ leading to α -alkoxy radical 18, which is finally trapped by hydride. When the reducing agent employed was tributyltin deuteride, the product was specifically labeled only at the methoxy group. This spectacular rearrangement¹² highlights the highly crowded topology of the taxol core and the tendency of the functionalities in the concave face of the molecule to strongly interact with each other. When the reaction mixture yielding **3** was carefully scrutinized for a similar

Scheme III



^a Drug concentration required to inhibit cell proliferation to 50% vs untreated cells (incubated at 37 °C for 72 h).

rearrangement product, none was found. Evidently, the C-10 acetate function acts as a deterrent with respect to the radical cascade. Although the nature of this effect is not obvious at this stage, this phenomenon is once again an illustration of the subtle interplay of the various functions in the molecule.

The cytotoxicity of 4 was essentially identical to that of taxol, while 12 was slightly less active. Detailed cytotoxicity data for compound 4 and 12 are listed in Table I.¹³ A more complete biological profile of these and related deoxygenated taxanes will be reported separately.

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Supplementary Material Available: Experimental details and ¹H-NMR spectra of key compounds (18 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.

⁽¹¹⁾ For the related epoxide fragmentation, see: Sabatino, E. C.; Gritter, R. J. J. Org. Chem. 1963, 28, 3437. For the fragmentation of cyclobutylcarbinyl radicals, see: Beckwith, A. L. J.; Moad, G. J. Chem. Soc., Perkin Trans. 1 1980, 1083.

⁽¹²⁾ For a review see: Beckwith, A. L. J.; Ingold, K. U. In *Free Radical Rearrangements*; De Mayo, P., Ed.; Academic Press: New York, 1980; Vol. 1, p 161.

⁽¹³⁾ The results are the average of three determinations. For details of the *in vitro* cytotoxicity assay: Scudiero, D. A.; Shoemaker, R. H.; Paull, K. D.; Monks, A.; Tierney, S.; Nofziger, T. H.; Currens, M. J.; Seniff, D.; Boyd, M. R. *Cancer Res.* 1988, 48, 4827.